

Dengue Virus Transmission via Deceased Renal Allograft: A Case Report Highlighting the Need for Donor Screening in Endemic Areas

Muhammad Abdul Mabood Khalil^{a, b} , Nihal Mohammed Sadagah^a, Alfatih Abdalla Altom^a,
Ahmed Abdelahad Basha^a, Hinda Hassan Khideer Mahmood^a,
Hisham Ismael Mohamed Sakran^a, Ibrahim Mohammed Nasser Assiri^a,
Ghaleb Anas Aboalsamh^a, Salem H. Al-Qurashi^a

Abstract

Dengue virus infection (DVI) has multiple routes of transmission. Modes of transmission include mosquito bites, perinatal transmission, blood transfusions, organ transplantation, needle stick injuries, or laboratory accidents. DVI in kidney transplant recipients is common in an endemic area. In an endemic area, it is usually caused by a mosquito bite. Solid organ transplantation, including the kidney, has been implicated in the transmission of DVI in the immediate post-transplant period. We describe a case of end-stage renal disease in which the patient got DVI immediately after getting a kidney from a deceased kidney donor. Our patient presented with fatigue and pain around the graft, anemia, thrombocytopenia, coagulopathy, hepatitis, and was found to have a hematoma around the graft. The recipient of the second kidney from the same donor also presented with fever and was found to have dengue. We describe our experience with managing our case, along with a detailed literature review of similar cases transmitted through renal allograft.

Keywords: Dengue virus infection; Transmission by renal allograft; Kidney transplant

Introduction

Dengue is an RNA virus transmitted primarily by the mosquito *Aedes aegypti* [1]. In endemic regions, dengue infection in kidney transplant recipients has mainly been attributed to bites from infected *Aedes aegypti* mosquitoes [2]. The disease may

range from mild to severe, causing morbidity [3] and mortality [4]. Although most dengue virus infections (DVI) are caused by direct bites from the mosquito *Aedes aegypti*, they can be rarely transmitted through solid organ transplantation of infected organs [5]. We share our experience with DVI transmitted by infected kidneys from a deceased donor and take the opportunity to do a literature review of the clinical presentation and outcome of DVI transmitted by infected renal allografts.

Case Report

Investigations

A 27-year-old female (recipient 1), who is known to have type 1 diabetes, hypertension, and end-stage renal disease on hemodialysis, underwent deceased kidney transplantation on January 28, 2023. The transplantation was ABO compatible, and both donors and recipients were blood group B positive. Human leukocyte antigen (HLA) typing showed three antigen matches. There were two donor-specific antibodies (DSAs) against donor HLA CW*(12:03) with a mean fluorescence intensity (MFI) of 798, and against HLA DRB5*01:01 (DR51) with an MFI of 709. Flow cross-match for T-cell immunoglobulin (Ig)G was negative, with a mean channel shift of -51. Flow cross-match for B-cell IgG was negative, with a mean channel shift of -48. The right kidney with a double ureter from a deceased donor was implanted in King Fahad Armed Forces Hospital on January 28, 2023, in the right iliac fossa. The recipient was treated with antithymocyte globulin (ATG), and a total dose of 4 mg/kg of ATG was administered. The recipient was started on prednisolone (Pred) and mycophenolate mofetil (MMF) as per protocol. Tacrolimus (TAC) was started on day 6. There were no medical or surgical complications. There was no delayed graft function. The patient was discharged on February 7, 2023, with a creatinine of 150 $\mu\text{mol/L}$. She presented on February 8, 2023, with pain over the graft and not feeling well. There was no fever, dysuria, or pyuria. The patient did not report any vomiting or loose motions. Clinically, she has pallor with tenderness around the graft area. Otherwise, she was afebrile with a respiratory rate of 16 breaths per minute,

Manuscript submitted May 17, 2025, accepted August 7, 2025
Published online August 22, 2025

^aCenter of Renal Diseases and Transplantation, King Fahad Armed Forces Hospital, Al Andalus, Jeddah 23311, Saudi Arabia

^bCorresponding Author: Muhammad Abdul Mabood Khalil, Center of Renal Diseases and Transplantation, King Fahad Armed Forces Hospital, Al Andalus, Jeddah 23311, Saudi Arabia. Email: doctorkhalil1975@hotmail.com

doi: <https://doi.org/10.14740/jmc5140>

blood pressure of 161/100 mm Hg, and oxygen saturation of 100% with a normal chest and cardiovascular examination.

Diagnosis

Lab workup showed a significant drop in hemoglobin from 14 g/dL to 6 g/dL. Her white blood cell (WBC) count increased from $5 \times 10^9/L$ to $16 \times 10^9/L$, and then to $30 \times 10^9/L$. She had a significant drop in her platelet count to $56 \times 10^9/L$. She had mild derangement of liver enzymes, with alanine aminotransferase (ALT) at 76 U/L, aspartate aminotransferase (AST) at 148 U/L, and alkaline phosphatase at 104 U/L. Her bilirubin was 7 $\mu\text{mol/L}$. She also had derangement of coagulation with a prothrombin time (PT) of 45 s, activated partial thromboplastin time (APTT) of 51.3 s, and an international normalized ratio (INR) of 3.79. Fibrinogen levels were measured three times and were normal. Blood cultures from the peripheral vein and right internal jugular catheter were negative. Urine culture was also negative. Von Willebrand antigen was high (286%). Von Willebrand ristocetin cofactor was normal (150%). Coagulation factor XIII was normal (107%).

Ultrasound (US) of the graft with Doppler showed normal perfusion with a collection around the graft. Patient underwent a non-enhanced computed tomography (CT) scan without contrast of the abdomen and pelvis. The transplanted kidney was seen in the right iliac fossa, measuring 10 cm. A significant, extensive, and heterogeneous soft tissue density structure was observed surrounding the transplanted kidney, occupying the right iliac fossa. It extended from the superior to the inferior liver edge and inferiorly to the right side of the urinary bladder, with further extension around the medial aspect of the transplanted kidney, measuring approximately $9 \times 11 \times 15$ cm in diameter, with a Hounsfield unit of 38. It demonstrated heterogeneous density along with fluid density and fluid levels. These findings suggested the presence of a hematoma around the graft.

The patient was re-explored on the same day of operation in the operating theatre, and no bleeding point was identified. The surgeon drained the hematoma and assumed that it was due to medical causes. Given thrombocytopenia, deranged liver enzymes, and coagulopathy, testing was done for dengue virus. The blood sample was analyzed using reverse transcription polymerase chain reaction (RT-PCR), which detected dengue virus. Furthermore, the patient tested positive for anti-dengue IgM antibodies, while IgG antibodies were reported negative. Although no pre-transplant serum or plasma samples were available to assess dengue immunity, the post-transplant positive non-structural protein 1 (NS1) antigen, IgM, and PCR, along with negative IgG, are consistent with a primary acute infection.

The second kidney was implanted in the recipient (recipient 2) at another hospital in the city. Upon the diagnosis of DVI in our case, the transplant team caring for the second recipient was also informed. Recipient 2 developed fever and body aches after 9 days of the transplant. The patient had mild derangement of liver enzymes with thrombocytopenia. Given the recipient 1's history of DVI, prompt screening was conducted

to rule out the infection. The patient was found to have dengue IgM antibodies and was NS1 antigen positive. However, IgG antibodies were reported negative. These results are consistent with a primary acute dengue infection. Although PCR and convalescent IgM testing were not performed, the serological profile supports recent infection rather than past exposure.

Although the detailed history from the other hospital was unavailable, the patient was treated with intravenous hydration and paracetamol and made a complete recovery. Since DVI is endemic in western Saudi Arabia, we assumed both recipients got DVI from the donor through transplanted kidneys.

We reviewed donor history in detail. The donor was a 23-year-old gentleman who was found semiconscious in the street and was brought to the hospital by ambulance on January 23, 2023. He has no past comorbidities. The patient was clinically unconscious with BP 117/68 mm Hg, pulse 102/min, and had a temperature of 37 °C. He had a temperature of 38 °C on January 26, 2023. His initial CT scan showed cerebral edema, and there was no evidence of infarct or hemorrhage. Magnetic resonance imaging (MRI) of the brain found cavernous sinus thrombosis. The CT scan of the chest, abdomen, and pelvis revealed a mild right pleural effusion with multiple alveolar infiltrates in both lungs, which became confluent posteriorly, forming consolidation with air bronchograms. His blood and urine cultures were negative. He has mild coagulopathy with thrombocytopenia along with deranged liver enzymes. The patient was admitted to the intensive care unit (ICU). Intubation was done, and he was ventilated. He was resuscitated with intravenous saline and was started on meropenem 500 mg every 8 h, along with vancomycin 1 g every 8 h. Despite respiratory support, antibiotics, and intravenous fluids, the patient remained comatose. Unfortunately, the patient did not show any neurological recovery and was diagnosed with brain death, and the family agreed to organ donation. We believe that the constellation of findings, including thrombocytopenia, derangement of liver enzymes, cavernous sinus thrombosis, and mild coagulopathy, were likely due to DVI.

Dengue is endemic in Saudi Arabia. Unfortunately, screening is not universally done in Saudi Arabia in deceased donors. Epidemiologically, the infected kidneys from the deceased donor implanted in both recipients were the likely source of transmission. Both recipients had no symptoms at baseline. They developed dengue between 9 and 10 days, which is consistent with the incubation period of 4 - 10 days of DVI. The recipients were in two different hospitals, and there was no epidemic or cluster at the time of DVI in either recipient. Most importantly, both recipients shared a common source: they had received kidneys from the same deceased donor.

Treatment

After evacuation of the hematoma, the patient received three units of packed red blood cells, four units of fresh-frozen plasmas, and two units of platelet concentrates. Her MMF was stopped, and she was treated with intravenous meropenem, given leukocytosis initially on an empirical basis. Upon diagnosis of DVI and negative culture, meropenem was held, and

the patient was resuscitated with intravenous saline and blood products. Subsequently, she received four pints of packed red blood cells, six platelet transfusions, and five doses of intravenous immunoglobulin (IVIG) at 0.4 g/kg during her stay. One week after admission, her WBC count was $5.22 \times 10^9/L$, hemoglobin was 9.7 g/dL, and platelets count was $55 \times 10^9/L$. Her liver enzymes normalized, with ALT at 18 U/L, AST at 33 U/L, and alkaline phosphatase at 71 U/L. Her clotting profile also normalized. However, her creatinine remained elevated at 175 $\mu\text{mol/L}$. The patient remained stable over the next 13 days.

Follow-up and outcomes

The patient's platelet count had normalized by February 20, 2023, to $185 \times 10^9/L$, with normalization of liver function test. She was discharged on February 28, 2023, with a creatinine level of 144 $\mu\text{mol/L}$. For the last 2 years since discharge, the patient has been under follow-up in the transplant clinic with stable graft function.

Discussion

The first dengue fever outbreak in Saudi Arabia was reported in 1994 in Jeddah, where 289 confirmed cases were reported. Since the first outbreak, there were numerous outbreaks in 2004, 2005, and 2006 [6]. Following these outbreaks, dengue is endemic in the cities of Jeddah and Makkah [6]. By the year 2009, the Ministry of Health in the Saudi Arabia had reported a total of 3,350 cases of dengue fever and a reported case fatality rate of 4.6 per thousand [7].

DVI has many manifestations, including thrombocytopenia, hepatitis, and coagulopathy [8]. The clinical findings, including a hematoma around the graft, thrombocytopenia, derangement of liver enzymes, and coagulopathy, led to suspicion of possible DVI, which was confirmed by RT-PCR in our case. We believe that the transplanted kidney was the source of infection, as the second recipient of the deceased donor's kidney also developed a fever and was found to have IgM antibodies against the dengue virus. Since the donor resided in an endemic area, transmission to both recipients via the renal allograft is a plausible explanation. The theoretical possibility of a primary infection of both recipients from the community simultaneously is less likely, as there was no epidemic. The possibility of thrombocytopenia, deranged liver enzymes, and coagulopathy [8] is an established manifestation of DVI. Thrombosis in various venous beds, including cerebral venous thrombosis, has been reported in the literature with DVI [9-11]. Fever, derangement of liver enzymes, mild coagulopathy, and cavernous sinus thrombosis in the deceased donor were possibly attributable to dengue infection. Dengue is endemic in Saudi Arabia. Unfortunately, screening is not universally done in Saudi Arabia in deceased donors. Both recipients from the same donor developed DVI at two separate centers. There was no local outbreak, making vector-borne transmission less likely.

The clinical presentation of DVI in kidney transplants differs from that of the general population. Fever, myalgia, arthralgia, and headache were significantly lower than in the normal population, while pleural effusions and ascites were observed more frequently [12]. Similarly, severe DVI is more common in transplant patients [12]. Like other viral illnesses, antimetabolites, including MMF and azathioprine, are typically withheld or reduced, especially in cases of leukopenia [13]. As our patient has severe DVI, MMF was withheld while TAC and Pred were continued. Care should be taken to restart the antimetabolites as soon as possible during recovery to minimize the risk of rejection.

Dengue transmission by solid organ transplantation (kidney, heart, and liver) has been reported [14]. Dengue has been implicated in glomerular injury [15]. Viral antigens have been detected in kidney tubular epithelial cells of infected individuals [16]; therefore, kidneys from deceased or live donors can potentially transmit infection to the recipient. Our literature search identified two cases from Brazil [5], two cases from France [17], two cases from Singapore [18, 19], one case from Thailand [20], and our present case. Summaries of all these case reports are shown in Table 1 [5, 17-20].

Approximately 78% (7/9) of transmissions occurred in deceased kidney donation as compared to 22% (2/9) in live kidney donation. This reflects that, unlike deceased kidney donation, live kidney donors are extensively evaluated in a well-planned manner, and sick donors are usually excluded or put on the waiting list until the acute issue is resolved. However, the history of DVI in a live kidney donor can still be a source of infection in a potential recipient [19]. Viral antigens are present in the tubular epithelium [16], and theoretically, the transmission of viral particles to the recipient and reactivation in the presence of intense immunosuppression may explain this phenomenon. The clinical course has been mild in 44.4% (4/9) of the cases, 33.3% have dengue hemorrhagic fever (3/9), and 22.2% (2/9) had dengue shock syndrome. Most of the instances of DVI transmitted by transplanted kidney presented 4 - 13 days post-transplantation [5, 18-20]. Derangement of liver enzymes was present in 88% of cases. This was followed by thrombocytopenia, which was present in 77.9% of the cases. One of the significant complications that we observed in 55.5% (5/9) of these patients was peri-graft hematoma around implanted kidneys [5, 17, 19, 20], which increased morbidity as these patients needed blood products and surgical evacuation of the collection. Though there was no mortality, the occurrence of DVI in the immediate post-transplant period led to more re-exploration [17, 18, 20] and a prolonged stay of 20 - 35 days [5, 17, 18]. Management of kidney transplant recipients includes symptomatic treatment, hydration, avoidance of nephrotoxic medications, and reduction of immunosuppression in the presence of cytopenia [21]. There are no specific guidelines for reducing immunosuppression. We withheld MMF and reintroduced it later once the patient fully recovered.

Some countries, such as France [22] and Singapore [23], have adopted policies for dengue virus screening before kidney donation. Important guidelines, such as those from Kidney Disease: Improving Global Outcomes (KDIGO) [24] and the American Society of Transplantation [25], provide clear recommendations for the evaluation of kidney donors with hepa-

Table 1. Summary of Case Reports Reporting DVI by Renal Allograft

Num-ber	Reference	Journal	Age/ gender	Type of renal transplantation	Clinical features & di- agnostic test	Immunosup- pression	Management	Outcome
Case 1	Rosso et al, 2018 [5]	Braz J Infect Dis	31/F	Deceased kidney transplantation	Needed readmission. Presented on day 8 with fever, vomiting, diarrhea, jaundice. Thrombocytopenia, lymphopenia, hepatitis. Pain over the graft with peri-renal hematoma of 1,300 mL	Pred, cyclosporine (CyA), MMF	Drainage of perirenal hematoma and transfusion of red blood cells, platelets, and fresh frozen plasma	Alive and was discharged on day 25
IgG+ IgM+								
Donor serum retrospec-tively checked and found to be NS1 antigen positive								
NS1+ RT-PCR+ (DEN4)								
Case 2	Rosso et al, 2018 [5]	Braz J Infect Dis	48/F	Deceased kidney transplantation	Presented on day 4 with fever and elevation of transaminases	Pred, CyA, MMF	Asymptomatic and conservative management. Patient was given oral ciprofloxacin for UTI.	Alive
Has E. coli UTI								
Donor serum was retrospec-tively checked and found to have NS1 antigen								
IgG- IgM+ NS1- PCR-								
Case 3	Lecadieu et al, 2021 [17]	Am J Trop Med Hyg	58/M	Deceased kidney transplantation	Has delayed graft function and presented on day 11 with hemodynamic instabil-ity (blood pressure 76/43 and heart rate 119, abdominal pain, peri-graft infected collection, anemia, thrombocytopenia, and deranged liver enzymes.	ATG, MP, MMF, TAC	Needed vasopressors, ventila-tion, and dialysis, and needed transfusion of platelet concentrate, packed red cells, and fresh frozen plasma. The course was compli-cated by an infected hematoma caused by <i>Staphylococcus epider-midis</i> (treated with vancomycin) and ventilator-associated pneu-monia by <i>Klebsiella pneumo-niae</i> , treated with meropenem	Alive and discharged on day 39 with creatinine 288 µmol/L
Blood PCR posi-tive for DENV-1								
Case 4	Lecadieu et al, 2021 [17]	Am J Trop Med Hyg	61/M	Deceased kidney transplantation	Presented on day 12 with thrombocytopenia (64 × 10 ⁹ /L) and deranged liver enzymes (AST 220 IU/L and ALT 181 IU/L) and a nor-mal clinical examination.	ATG, MP, MMF, TAC	Treated symptomatically	Discharged home on day 35 with creatinine 271 µmol/L
Both blood and urine PCR were positive for DENV-1, and den-tigue serology was negative (IgM index 0.5 and IgG index 0.1).								
Case 5	Sim et al, 2021 [18]	Am J Transplant	63/M	Deceased kidney transplantation	Presented on day 5 with fever, thrombocytopenia, and deranged liver enzymes. No delayed graft functions	Induction basiliximab		Recovered uneventfully and was dis-charged home

Table 1. Summary of Case Reports Reporting DVI by Renal Allograft - (continued)

Num-ber	Reference	Journal	Age/ gender	Type of renal transplantation	Clinical features & di- agnostic test	Immunosup- pression	Management	Outcome
Case 6	Sim et al, 2021 [18]	Am J Transplant	39/M	Donor negative by blood RT-PCR. However, urine RT-PCR positive	RT-PCR detected	Maintenance TAC, Pred, and MMF. No comment about modifica- tion of immu- nosuppression		
				Deceased kidney transplantation	Asymptomatic with normal platelet count and mildly high alanine transaminase	Induction with ATG and MP		Recovered uneventfully and was dis- charged home.
Case 7	Tan et al, 2005 [19]	Nephrol Dial Transplant	23/M	Live kidney trans- plantation	Presented on day 5 with fever, gastrointestinal bleeding, thrombocytopenia, leukopenia, right effusion, hemorrhagic ascites, and a large retroperito- neal hematoma arising from the bed of the transplanted kidney	Maintenance with TAC, Pred, and MMF. No comment about the modification of immunosup- pression		
				A donor had dengue fever 6 months back	RT-PCR detected	Induction MP	His MMF was stopped; packed red cell and platelet transfu- sions were given; granulocyte colony-stimulating factor given; hematoma drained surgically.	Full recovery and graft func- tion remained excellent
Case 8	Tangnara- ratchakit et al, 2012 [20]	Transplant Proc	15/F	Live kidney trans- plantation	Presented with perinephric hematoma, pancytopenia, ascites, effusion, and shock	Induction with MP followed by Pred, cyclo- sporine, MMF	Patient was treated via surgi- cal drainage, transfusions, and hemodynamic support	Patient sur- vived with full recovery of graft function
Case 9	Present case 1		27/F	Deceased kidney transplantation	Presented on day 11 with fatigue, hematoma, anemia, thrombocytopenia, deranged liv- er enzymes, and coagulopathy	Induction with ATG	MMF was held; IV fluid given; packed red cells, plate- lets, and FFP were given; hematoma was drained.	Discharged on day 20 with creatinine of 144 µmol/L

DVI: dengue virus infection; F: female; M: male; Pred: prednisolone; MMF: mycophenolate mofetil; TAC: tacrolimus; ALT: alanine aminotransferase; AST: aspartate aminotransferase; MP: methylprednisolone; UTI: urinary tract infection; Ig: immunoglobulin; ATG: antithymocyte globulin; IV intravenous; *E. coli*: *Escherichia coli*; RT-PCR: reverse transcription polymerase chain reaction; NS1: non-structural protein 1; DENV-1: dengue virus serotype 1.

titis B virus, hepatitis C virus, and human immunodeficiency virus. However, these guidelines do not recommend screening for the dengue virus in kidney donors before donation. It is essential to screen for DVI at least in endemic areas [21]. Given the variability in current practices, international transplant societies need to establish expert consensus to ensure uniform practice and provide guidance for screening policies worldwide. There are no clear guidelines on whether kidneys from a donor infected with dengue should be used. In a center in Brazil, where dengue is endemic, solid organ transplant donors who were NSI antigen positive were excluded. Those with negative NSI antigen and having positive antibodies were allowed to donate. Authors reported no DVI after adopting this strategy [5]. Checking PCR for the presence of viremia is another way to predict transmission. If viremia is detected, the infectious reservoir will be high, and the risk of transmission will be high [26]. There is also a need for international transplant societies to develop consensus on the utilization of kidneys obtained from deceased donors. Therefore, if the dengue virus is detected by serological test in a deceased donor, checking for active viremia using PCR may be applicable for assessing the risk of transmission. Another critical point to remember is that if a recipient has a previous infection, a subsequent infection with a different dengue virus may have a worse outcome [27]. Therefore, it is essential to inquire about the potential recipient's history of DVI. It is also necessary to explain to the recipient of a kidney from an infected donor the risk of transmission, bleeding, peri-graft hematoma, and prolonged hospital stay.

We acknowledge a few limitations. As a single case report, the findings cannot be generalized to other transplant recipients. Since there is no protocol for screening deceased kidney donors in Saudi Arabia, it becomes difficult to ascertain the donor's infection status definitively and assess the risk of transmission. Another limitation of this report is the unavailability of the detailed clinical history of the second kidney recipient, who was treated at another institution.

Our case report has a few strengths. This is the first case report from Saudi Arabia where the virus is endemic, identifying an important question to screen for the dengue virus in deceased organ donation. Secondly, an extensive review of DVI in renal allograft recipients provides essential information about the course of the disease post-implantation. It identifies gaps in recommendations from various organizations and societies to guide screening for DVI in deceased donors in endemic areas.

Learning points

This case, along with the available literature, demonstrates that dengue virus transmission through renal allografts is possible. The post-transplant clinical course can be highly variable. This can range from mild dengue viral illness to severe dengue hemorrhagic fever. Screening of deceased donors in endemic regions is essential. This will help to prevent transmission to transplant recipients. There is a clear need for transplant societies and organizations to develop standardized guidelines for dengue screening in both deceased and live donors. The

potential risks of transmission and associated complications should be thoroughly discussed with recipients during the consent process. Management of immunosuppression during active dengue infection also remains an area of uncertainty. Therefore, a review of current evidence and the development of consensus guidelines on adjustments of immunosuppression are needed.

Acknowledgments

None to declare.

Financial Disclosure

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflict of Interest

The authors declare that they have no competing interests.

Informed Consent

Written informed consent was obtained from the patient.

Author Contributions

MAMK, SHA, NMS, and GAA conceived the study idea. MAMK drafted the initial version, and all authors (NMS, AAA, MAMK, AAB, HHKM, HIMS, IMNA, GAA, SHA) critically reviewed it. MAMK revised and all authors (NMS, AAA, MAMK, AAB, HHKM, HIMS, IMNA, GAA, SHA) approved the final manuscript.

Data Availability

All data in our report were obtained from the patient's hospitalization. Any inquiries regarding the supporting data availability of this study should be directed to the corresponding author.

Abbreviations

DVI: dengue virus infection; MMF: mycophenolate mofetil; TAC: tacrolimus; Pred: prednisolone

References

1. Humayoun MA, Waseem T, Jawa AA, Hashmi MS, Akram J. Multiple dengue serotypes and high frequency

- of dengue hemorrhagic fever at two tertiary care hospitals in Lahore during the 2008 dengue virus outbreak in Punjab, Pakistan. *Int J Infect Dis.* 2010;14(Suppl 3):e54-59. [doi pubmed](#)
2. Nasim A, Anis S, Baqi S, Akhtar SF, Baig-Ansari N. Clinical presentation and outcome of dengue viral infection in live-related renal transplant recipients in Karachi, Pakistan. *Transpl Infect Dis.* 2013;15(5):516-525. [doi pubmed](#)
 3. Costa SD, da Silva GB, Jr., Jacinto CN, Martiniano LV, Amaral YS, Paes FJ, De Mattos Brito Oliveira Sales ML, et al. Dengue fever among renal transplant recipients: a series of 10 cases in a tropical country. *Am J Trop Med Hyg.* 2015;93(2):394-396. [doi pubmed](#)
 4. Prasad N, Bhadauria D, Sharma RK, Gupta A, Kaul A, Srivastava A. Dengue virus infection in renal allograft recipients: a case series during 2010 outbreak. *Transpl Infect Dis.* 2012;14(2):163-168. [doi pubmed](#)
 5. Rosso F, Pineda JC, Sanz AM, Cedano JA, Caicedo LA. Transmission of dengue virus from deceased donors to solid organ transplant recipients: case report and literature review. *Braz J Infect Dis.* 2018;22(1):63-69. [doi pubmed](#)
 6. Alhaeli A, Bahkali S, Ali A, Househ MS, El-Metwally AA. The epidemiology of Dengue fever in Saudi Arabia: A systematic review. *J Infect Public Health.* 2016;9(2):117-124. [doi pubmed](#)
 7. Ministry of Health, Department of Statistics Health statistical year book 2009 Saudi Ministry of Health, Riyadh, KSA. 2010. ISSN: 1319-3228.
 8. Khalil MA, Sarwar S, Chaudry MA, Maqbool B, Khalil Z, Tan J, Yaqub S, et al. Acute kidney injury in dengue virus infection. *Clin Kidney J.* 2012;5(5):390-394. [doi pubmed](#)
 9. Vasanthi N, Vairamon PM, Gowtham T, Das AK. Unusual presentation of dengue fever-cerebral venous thrombosis. *J Clin Diagn Res.* 2015;9(6):OD09-10. [doi pubmed](#)
 10. Hameed S, Hashmat O, Memon M, Wasay M. Cerebral venous sinus thrombosis with positive Dengue serology. *PJNS.* 2019;14(5):41-43.
 11. da Costa PS, Ribeiro GM, Junior CS, da Costa Campos L. Severe thrombotic events associated with dengue fever, Brazil. *Am J Trop Med Hyg.* 2012;87(4):741-742. [doi pubmed](#)
 12. Weerakkody RM, Patrick JA, Sheriff MH. Dengue fever in renal transplant patients: a systematic review of literature. *BMC Nephrol.* 2017;18(1):15. [doi pubmed](#)
 13. Tan SSX, Ho QY, Thangaraju S, Tan TT, Kee T, Chung SJ. Dengue virus infection among renal transplant recipients in Singapore: a 15-year, single-centre retrospective review. *Singapore Med J.* 2024;65(4):235-241. [doi pubmed](#)
 14. Cedano JA, Mora BL, Parra-Lara LG, Manzano-Nunez R, Rosso F. A scoping review of transmission of dengue virus from donors to recipients after solid organ transplantation. *Trans R Soc Trop Med Hyg.* 2019;113(8):431-436. [doi pubmed](#)
 15. Boonpucknavig V, Bhamarapravati N, Boonpucknavig S, Futrakul P, Tanpaichitr P. Glomerular changes in dengue hemorrhagic fever. *Arch Pathol Lab Med.* 1976;100(4):206-212. [pubmed](#)
 16. Jessie K, Fong MY, Devi S, Lam SK, Wong KT. Localization of dengue virus in naturally infected human tissues, by immunohistochemistry and in situ hybridization. *J Infect Dis.* 2004;189(8):1411-1418. [doi pubmed](#)
 17. Lecadiou A, Teyseyre L, Larsen K, Vidal C, Caron M, Traversier N, Ludovic Di a, et al. Case report: transmission of dengue virus from a deceased donor to a kidney transplant recipient previously infected by dengue virus. *Am J Trop Med Hyg.* 2021;104(6):2199-2201. [doi pubmed](#)
 18. Sim JXY, Gan ES, Tan HC, Choy MM, Wong HM, Tan BH, Kee T, et al. Aviremic organ transplant dengue virus transmission - A case report. *Am J Transplant.* 2021;21(5):1944-1947. [doi pubmed](#)
 19. Tan FL, Loh DL, Prabhakaran K, Tambyah PA, Yap HK. Dengue haemorrhagic fever after living donor renal transplantation. *Nephrol Dial Transplant.* 2005;20(2):447-448. [doi pubmed](#)
 20. Tangnaratchakit K, Tirapanich W, Tapaneya-Olarn W, Sumethkul V, Sirachainan N, Watcharananan S, Leenanunpunth C, et al. Severe nonfebrile dengue infection in an adolescent after postoperative kidney transplantation: a case report. *Transplant Proc.* 2012;44(1):303-306. [doi pubmed](#)
 21. Delfino VDA, Mazzali M. Dengue in kidney transplanted patients: additions to the puzzle! *J Bras Nefrol.* 2022;44(1):6-8. [doi pubmed](#)
 22. Haut Conseil de la sante publique (France). Avis du 19 decembre 2024 relatif a la securisation des produits du corps humain, notamment des greffons, contre le virus de la dengue. Paris: HCSP. 2024 [cited Aug 4, 2025]. Available from: <https://www.hcsp.fr/Explore.cgi/AvisRapports>.
 23. Tan SSX, Nordin SB, Tan CK, Tan TT, Chung SJ, Chan KS, Tan BH. Donor-derived dengue infections - a review of screening protocol and outcomes in an endemic country. *Transpl Infect Dis.* 2024;26(Suppl 1):e14356. [doi pubmed](#)
 24. Lentine KL, Kasiske BL, Levey AS, Adams PL, Alberu J, Bakr MA, Gallon L, et al. KDIGO clinical practice guideline on the evaluation and care of living kidney donors. *Transplantation.* 2017;101(8S):S1-S109. [doi pubmed](#)
 25. Wolfe CR, Ison MG, Practice ASTIDCo. Donor-derived infections: guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. *Clin Transplant.* 2019;33(9):e13547. [doi pubmed](#)
 26. Matangkasombut P, Manopwisedjaroen K, Pitabut N, Thaloengsok S, Suraamornkul S, Yingtaeesak T, Duong V, et al. Dengue viremia kinetics in asymptomatic and symptomatic infection. *Int J Infect Dis.* 2020;101:90-97. [doi pubmed](#)
 27. Shih HI, Wang YC, Wang YP, Chi CY, Chien YW. Risk of severe dengue during secondary infection: A population-based cohort study in Taiwan. *J Microbiol Immunol Infect.* 2024;57(5):730-738. [doi pubmed](#)